

## Colorimetric Microplate Assay for 2-Thiobarbituric Acid Reactive Substances (TBARS)

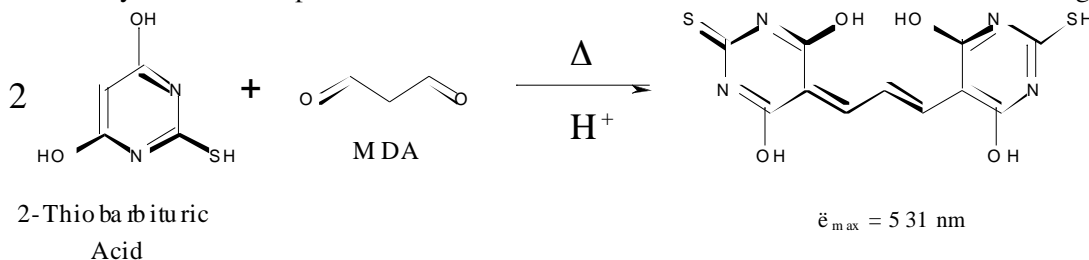
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### INTRODUCTION

2-ThioBarbituric Acid Reactive Substances (TBARS) are naturally present in biological specimens and include lipid hydroperoxides and aldehydes which increase in concentration as a response to oxidative stress.<sup>1,2</sup> TBARS assay values are usually reported in malonaldehyde (malondialdehyde, MDA) equivalents, a compound that results from the decomposition of polyunsaturated fatty acid lipid peroxides. The TBARS assay is a well-recognized, established method for quantifying these lipid peroxides, although it has been criticized for its reactivity towards other compounds other than MDA.<sup>3</sup> This kit contains modifications from the traditional TBARS assay to reduce background and the generation of TBA reactive substances to give a straightforward, reproducible and consistent method for analyzing biological samples for lipid peroxidation products.

### PRINCIPLES OF PROCEDURE

This assay is based on the reaction of a chromogenic reagent, 2-thiobarbituric acid, with MDA at 25°C. One molecule of MDA reacts with 2 molecules of 2-thiobarbituric acid *via* a Knoevenagel-type condensation to yield a chromophore with absorbance maximum at 532 nm, as shown below in Figure 1:



**Figure 1.** Reaction between 2-thiobarbituric acid and MDA under acidic conditions.

### MATERIALS PROVIDED

Component	Description	Amount	Storage	Cat. No.
<b>Indicator</b>	2-Thiobarbituric Acid	2 x 0.5 g	4°C	FR40a
<b>Acid Reagent</b>	10% Acid Solution in Dimethylsulfoxide	40 mL	4°C	FR40b
<b>MDA Standard</b>	10 mM Malonaldehyde Tetrabutylammonium Salt	100 $\mu$ L	4°C	FR40c
<b>Microplate</b>	96-well Microplate	2 plates	4°C	FR40d

### MATERIALS NEEDED BUT NOT PROVIDED

1. Spectrophotometric plate reader capable of reading at 532 nm (a 540 nm filter may also be used)
2. Polypropylene microcentrifuge tubes
3. Deionized water ( $dH_2O$ )
4. Adjustable micropipettes (10 – 1,000  $\mu$ L) and tips

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**STORAGE**

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1. The reagents are stable until the indicated kit expiration date if handled and stored properly.
2. When not in use, store the kit at 4°C for up to one year.
3. MDA standards should be used within 24 hours of preparation.
4. The Indicator Solution (combined 2-TBA and Acid Solution) can be stored at 4°C for one week.

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**WARNINGS AND PRECAUTIONS**

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1. Use aseptic techniques when opening and dispensing reagents.
2. Wear gloves and safety glasses when performing this assay, as the acid used is corrosive.
3. In case of accidental exposure to 2-TBA or Acid Reagent, thoroughly wash the exposed area with soap and water.
4. This kit is designed to work properly as provided and instructed. Additions, deletions or substitutions to the procedure or reagents are not recommended, as they may be detrimental to the assay.

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**PROCEDURAL NOTES**

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1. Do not leave the reagent bottles open. Replace the caps as soon as the desired volume is removed.
2. To minimize error due to handling, wipe the exterior bottom of the microplate wells with a lint-free paper towel.
3. **Sample Blanks:** It is HIGHLY recommended that, for each sample analyzed, a sample blank is also performed to account for background interferences. While Oxford's TBARS method has relatively low background, some background interferences are inherent in biological samples and cannot be avoided. The sample blank should contain the sample plus the Acid Reagent alone without TBA indicator. Following this procedure will result in more accurate, precise and reliable TBARS measurements.
4. There are sufficient standards and reagents to test 40 samples and 40 sample blanks in duplicate. Samples and sample blanks should be run together on the same plate using the same standard curve.

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**REAGENT PREPARATION**

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1. **Acid Reagent:** This solution may need to be thawed upon removal from 4°C. Allow to sit at room temperature for one hour. It is ready to use once thawed.
2. **Indicator Solution:** Add 10 mL of the Acid Reagent to the powdered contents of one vial of Indicator and shake until completely dissolved. One vial is sufficient for standards and 20 samples.
3. **20 µM MDA Standard Stock:** Dilute the 10 mM MDA Standard 1:500 in dH<sub>2</sub>O by adding 20 µL of 10 mM MDA to 9.98 mL dH<sub>2</sub>O. Prepare immediately prior to use.

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**SAMPLE STABILITY**

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Studies at OBR show that this kit provides optimum results with urine that is less than 48 hours old, and is preferably run immediately after sample collection. If this is not possible, samples should be frozen at -70°C to prevent loss of MDA and HAE<sup>4</sup> and sample oxidation. Samples should **not** be stored at -20°C. Samples should not be refrozen and should be protected from light to avoid photooxidation.

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**SAMPLE PREPARATION**

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When working with plasma, the sample should be deproteinated with an acid. Centrifuge and use the supernatant to perform the assay. This solution may appear cloudy after the reaction, and can be clarified by passing through a 0.2 µ syringe filter.

When working with urine, colored compounds contribute to the signal measured at 532 nm. This interference can be removed by running a sample blank with each sample.

## Urine

1. Urine samples can be used directly and should be assayed immediately. If the assay is to be performed on a different day, the sample should be stored at  $-70^{\circ}\text{C}$ .

## Plasma and Serum

1. Collect blood samples and process immediately per the collection tube instructions.
2. Prepare a saturated solution of ammonium sulfate.
3. Add  $100\ \mu\text{L}$  of saturated ammonium sulfate to  $0.5\ \text{mL}$  of serum or plasma in a test tube or microcentrifuge tube.
4. Add  $35\ \text{mg}$  TCA (trichloroacetic acid) to each sample and vortex. A cloudy precipitate should form.
5. Centrifuge the tubes and transfer the supernatant to a clean tube. Plasma and serum samples can be run without dilution. Samples are now ready for assay.

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## STANDARD CURVE PREPARATION

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Malondialdehyde is provided as a solution of the malondialdehyde tetrabutylammonium (MDA-TBA) salt in a slightly basic buffer because MDA itself is not stable. When mixed with the acidic Indicator Solution, the MDA-TBA molecule is acidified and generates MDA quantitatively.

Please see the **Reagent Preparation** section for preparing the  $20\ \mu\text{M}$  MDA Standard Stock.

**Table 1:** Colorimetric Standard Curve Preparation

Standard	MDA Conc. ( $\mu\text{M}$ )	Vol. of $\text{dH}_2\text{O}$ ( $\mu\text{L}$ )	Vol. of $20\ \mu\text{M}$ MDA Stock ( $\mu\text{L}$ )
S <sub>0</sub>	0	400	-
S <sub>1</sub>	0.5	390	10
S <sub>2</sub>	1.0	380	20
S <sub>3</sub>	2.5	350	50
S <sub>4</sub>	5.0	300	100
S <sub>5</sub>	10.0	200	200
S <sub>6</sub>	15.0	100	300
S <sub>7</sub>	20.0	-	400

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## ASSAY PROCEDURE

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### Free MDA

1. **Preparation of Standards and Samples:** Add each of the following reagents into microcentrifuge tubes and mix well.
  - **Standards:**  $200\ \mu\text{L}$  of standard and  $200\ \mu\text{L}$  of Indicator Solution.
  - **Samples:**  $200\ \mu\text{L}$  of sample and  $200\ \mu\text{L}$  of Indicator Solution.
  - **Blanks:**  $200\ \mu\text{L}$  of sample and  $200\ \mu\text{L}$  of Acid Reagent.
2. Incubate the Standards, Samples and Blanks at room temperature for 45 minutes.
3. Transfer  $150\ \mu\text{L}$  of each solution to the microplate and read at  $540\ \text{nm}$ . Please note that the optimum absorbance wavelength for this assay is  $532\ \text{nm}$ . See Scheme 1 on the following page for a sample plate layout.

### Total MDA

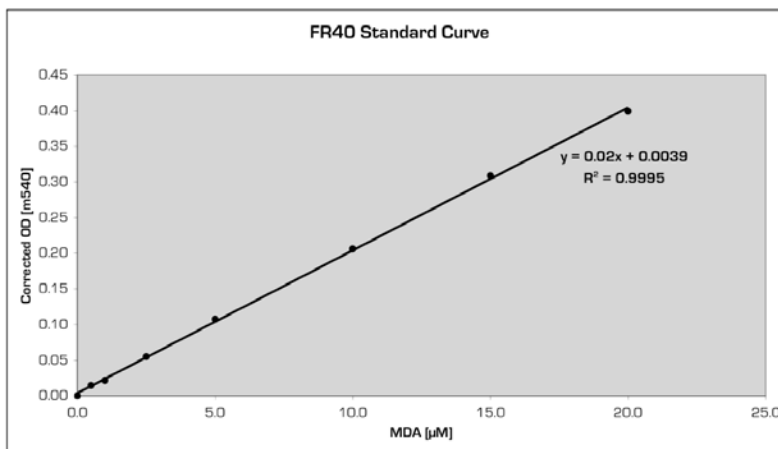
1. Prepare samples and standards exactly as above, but heat sample at  $65^{\circ}\text{C}$  for 45 minutes, then follow step 3 as above.

**Scheme 1: Sample Plate Layout**

	1	2	3	4	5	6	7	8	9	10	11	12
<b>A</b>	S0	S0	SPL1	SPL1	SPL5	SPL5	SPL9	SPL9	SPL13	SPL13	SPL17	SPL17
<b>B</b>	S1	S1	SB1	SB1	SB5	SB5	SB9	SB9	SB13	SB13	SB17	SB17
<b>C</b>	S2	S2	SPL2	SPL2	SPL6	SPL6	SPL10	SPL10	SPL14	SPL14	SPL18	SPL18
<b>D</b>	S3	S3	SB2	SB2	SB6	SB6	SB10	SB10	SB14	SB14	SB18	SB18
<b>E</b>	S4	S4	SPL3	SPL3	SPL7	SPL7	SPL11	SPL11	SPL15	SPL15	SPL19	SPL19
<b>F</b>	S5	S5	SB3	SB3	SB7	SB7	SB11	SB11	SB15	SB15	SB19	SB19
<b>G</b>	S6	S6	SPL4	SPL4	SPL8	SPL8	SPL12	SPL12	SPL16	SPL16	SPL20	SPL20
<b>H</b>	S7	S7	SB4	SB4	SB8	SB8	SB12	SB12	SB16	SB16	SB20	SB20

**CALCULATIONS**

1. Average the OD ( $A_{532}$ ) values obtained for all duplicated wells.
2. Plot a standard curve using the  $A_{532}$  OD value for each Standard versus the MDA concentration for each Standard. The equation of the line can be found using a linear fit method.
3. Subtract the OD ( $A_{532}$ ) for each Sample Blank from the Sample OD ( $A_{532}$ ) to obtain a Net OD ( $A_{532}$ ).
4. Calculate the MDA concentration for each Sample using the Net OD ( $A_{532}$ ) value and the equation generated by the Standard Curve. If the Samples were diluted, the result must be multiplied by the dilution factor. If you are testing serum or plasma samples and have followed the deproteinization step above, your dilution factor is 1.2.

**Figure 2: Typical Standard Curve****LIMIT OF DETECTION**

The limit of detection for the colorimetric assay has been determined to be 1.0  $\mu\text{M}$ .

**PERFORMANCE LIMITATIONS**

1. Although the standards in this assay will usually appear water clear, the samples may become colored. This is due to the formation of additional chromophores that absorb at various wavelengths other than 532 nm and will usually not interfere with the  $A_{532}$  signal.

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